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Cell Therapeutics, Inc.

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COMPOUNDS HAVING SELECTIVE HYDROLYTIC POTENTIALS

5 **Cross-Reference to Related Application**

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This application is a Continuation-in-Part Application of U.S. Applications Serial Nos. 08/199,368 and 08/306,091, filed February 18, 1994 and September 14, 1994, respectively.

10 **Technical Field of the Invention**

The invention provides compounds having selective (programmed) stability in adverse chemical and dynamic biologic environments. The disclosed compounds permit selective control of the extent and rate of conversion of the inventive compounds to a hydroxyl-substituted chiral xanthine compound.

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Background of the Invention

Drugs possessing a chiral center generally are developed, tested, manufactured and marketed as racemates, containing equivalent amounts of each respective enantiomer. In biologic systems, one enantiomer may be an active compound while the other enantiomer may have little or no effect. Therefore in some instances, skilled artisans may seek to enhance efficacy, improve potency or prolong therapeutic potential by isolating efficacious enantiomers from nonactive ones.

Enantiomerically pure compounds have been prepared by first synthesizing a racemate, and then, using chiral separation techniques, isolating the enantiomers. Manufacturing and dosing drugs as racemic mixtures means that each drug dose contains an equivalent amount of a corresponding isomer, sometimes having little or no therapeutic potential and quite possibly may cause unsuspected and undesirable side affects.

Thus, in many biologic systems, to attain, *inter alia*, greater potency or prolonged efficacy of compounds in the laboratory, a need exists to administer substantially pure enantiomers for use as pharmaceuticals. An example of one such pure enantiomer is the compound lisofylline, 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine, (described in U.S. Application Serial No. 08/307,554 filed on September 16, 1994 (now pending), the entire disclosure of which is incorporated by reference herein). Lisofylline is useful for a wide variety of therapeutic indications, including, for example, treating sepsis and sepsis syndrome, preventing multi-organ dysfunction associated with trauma, increasing the production of multilineage hematopoietic cells after cytoreductive therapies, promoting trilineage

engraftment after bone marrow transplantation, and alleviating the toxic side effect of interleukin-2 (IL-2), amphotericin B, cyclosporin A, FK506 or granulocyte macrophage colony stimulating factor (GM-CSF) therapies.

5 *In vitro* and *in vivo* studies and human clinical trials with i.v. administered lisofylline confirm therapeutic efficacy of this substantially pure enantiomer. Further studies in predictive animal models (*e.g.*, mouse, rat and dog models) using lisofylline as an oral formulation conclusively indicate that the chiral lisofylline is absorbed as a chiral molecule. Oral administration of a drug is often desired for, *inter alia*, ease of use, self-administered therapy and cost reduction; and thus, an orally administered drug, which eliminates more invasive i.v. 10 and i.p. procedures is desirable.

15 In general, therapeutic deficiencies of new or existing drugs usually result from difficulties in any of three phases of drug action: pharmaceutic, biopharmaceutic, and pharmacodynamic. The pharmaceutic phase refers to the chemical and physical environment of a drug prior to its absorption into the living system and includes the dosage form of the drug as well as tissues encountered at the site of administration. The biopharmaceutic phase of drug action include absorption, distribution, biotransformation, and elimination of the drug. The pharmacodynamic phase deals with the drug at its pharmacologic site(s) of action. Difficulty associated with any of these three phases of drug action can result in either a subtherapeutic or toxic response of the patient to drug therapy. Riley, T. N., "The Prodrug Concept and New 20 Drug Design and Development," *Journal of Chemical Education*, Vol. 65, No. 11, November pages 947-953. 1988.

25 In orally administered, enantiomerically pure pharmaceuticals, a compound must retain its efficacious properties and chemical and chiral structure throughout various degradative biologic systems enroute to the bloodstream after ingestion. Such systems, which are encountered enroute to merging with systemic blood flow include, for example, digestion (stomach gastric juices), metabolism (intestinal membrane, portal vein and liver blood) and blood filtration (kidney and especially liver). Skilled artisans strive to minimize a first pass effect. The first pass effect is defined as those processes, such as digestion, metabolism and blood filtration which alter the drug such that it becomes inactive. An objective in minimizing 30 the first pass effect is to increase bioavailability of orally administered pharmaceutical compounds. An increased systemic concentration of a therapeutic compound for a longer period of time corresponds to higher bioavailability.

35 A prodrug is defined as a pharmacologic precursor (parent) to an active compound, which itself alone may possess efficacious properties. Conversion of a prodrug to an active compound occurs by either chemical or enzymatic processes after administration. Riley, *supra*.

Altering the chemical structure of a compound may include modification of structural features requiring sophisticated chemical or biochemical mechanisms to generate the parent specie.

The preparation of a "prodrug" form of another compound can have a number of beneficial effects in terms of increasing therapeutic utility of the therapeutic compound. Such beneficial effects include, for example, improved chemical stability characteristics, improved absorption characteristics following oral administration, better taste to improve patient compliance upon oral administration, and improved pharmacodynamics and pharmacokinetic characteristics, including altering metabolism of the parent chiral drug.

Target compounds as prodrug candidates must be stable in a variety of chemical and biologic systems, including the gastrointestinal tract and whole blood so as to have maximum effect on increasing bioavailability of an intended pharmaceutical *in vivo*. Yet, these candidates need not liberate a parent compound quickly or completely prior to or during ingestion or in the liver to achieve this result, and often it is undesirable to do so.

Because of the complexity of some biologic systems and the specificity of compounds as they relate to particular chemical or enzymatic pathways, success of one prodrug approach is not conclusive of proof of success for other drugs having differing structures and targeted therapeutic potential. Problems with solubility, toxicity, stability, bioavailability all vary between drugs. Bundgaard *et al.*, "A Novel Solution-Stable, Water-Soluble Prodrug Type for Drugs Containing a Hydroxyl or an NH-Acidic Group," Journal of Medicinal Chemistry, Vol. 32, No. 12, December 1989.

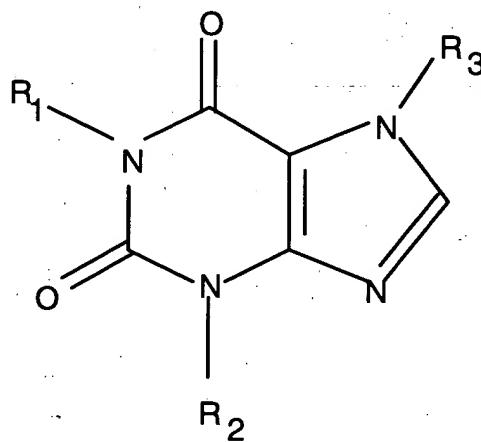
Hussain *et al.* disclose selected prodrugs of Naltrexone, used in treating opioid addiction. "Improvement of the Oral Bioavailability of Naltrexone in Dogs: A Prodrug Approach," Journal of Pharmaceutical Sciences, Vol. 76, No. 5, May 1987. The disclosed prodrugs include anthranilate, acetylsalicylate, benzoate and pivalate esters of Naltrexone. Naltrexone has numerous chiral centers, but Hussain *et al.* were concerned only with bioavailability of the racemic compound. Of the four esters disclosed, only the anthranilate and acetylsalicylate esters exhibited substantially higher bioavailability when compared with the bioavailability for orally administered Naltrexone. But, as acknowledged by Hussain *et al.*, there is no correlation between bioavailability and the *in vitro* hydrolysis half-life. Thus, identifying and isolating a prodrug, having the necessary characteristics to minimize or eliminate the first pass effect is a difficult task. In particular, lisofylline, its efficacious properties primarily a result of its specific enantiomeric stereochemistry, made a search for a corresponding oral prodrug of lisofylline very difficult.

The invention is in response to an identified need to develop compounds that have selective stability in adverse chemical and metabolic systems. Such compounds would ideally be capable of maintaining enantiomeric stereochemistry, but when exposed to an adverse

chemical or biologic environment (whether *in vitro* or *in vivo*), would maintain their chemical properties, having their own efficacious properties, or could be converted to a corresponding therapeutic compound. The invention results from these efforts to discover compounds, heretofore unknown, which structurally would have not only specific, variable abilities to maintain enantiomeric chemistry, but also programmed stability in dynamic, metabolic systems.

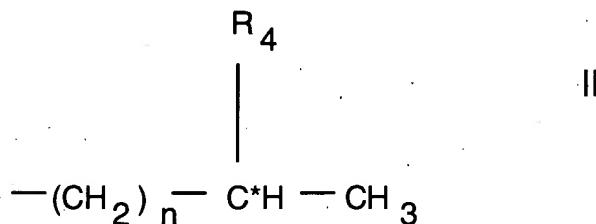
Summary of the Invention

The inventive compounds possess varying stabilities and different rates of hydrolysis to a corresponding alcohol. Such differences correlate to programmed hydrolytic potential. These differences are characteristic of varying chemical structures and thus programmed rates of hydrolysis are compound specific. More particularly, the invention provides a compound comprising a xanthine core of the formula I or a pharmaceutically acceptable salt, solvate or hydrate thereof:



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One of R₁ or R₂ is independently an aliphatic hydrocarbon having the formula II:



In formula II, n is an integer from about four to about eight, R₁ or R₂, which is other than

20 formula II, and R₃ are independently C₍₁₋₁₂₎ alkyl and C* is a chiral carbon atom. R₄ is an amino acid or carbohydrate attached to the chiral carbon atom C* via an ester linkage, or -O-X-(R₅)_m.

If R₄ is -O-X-(R₅)_m, m is two or three, X is selected from the group consisting of C, P or S and R₅ is a hydrogen atom; hydroxyl group; =O; substituted or unsubstituted C(1-10) alkyl, C(1-10) alkenyl, C(1-10) alkynyl, C(1-10) alkoxyl, C(1-10) oxoalkyl, or C(1-10) acetoxy, C(1-10) carboxyalkyl, or C(1-10) hydroxyalkyl group; -OR₆, R₆ being a substituted or unsubstituted C(1-10) alkyl, C(1-10) alkenyl, C(1-10) alkynyl; C(1-10) oxoalkyl; substituted or unsubstituted cyclic or heterocyclic group having from one- to three-rings, each ring containing from four to seven atoms.

These chiral inventive compounds provide for selectively stable compounds, both *in vitro* and *in vivo*. In more preferred compounds of the invention, a specific moiety chirally linked to the lisofylline skeleton permits control of the extent and rate of conversion of the inventive compounds to a corresponding alcohol. An acidic or basic environment, or when administered orally, various intestinal enzymes, or plasma or liver enzymes, hydrolyze the inventive compounds to form a corresponding alcohol. Thus, the present invention provides, *inter alia*, prodrugs for active chiral secondary ω-1 hydroxyl-substituted xanthine compounds.

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Brief Description of Drawings

Figure 1 is a schematic representation of a general synthesis of an inventive compound from lisofylline.

20 Figure 2 illustrates the amount of hydrolysis of various prodrug compounds with whole human blood.

Figures 3 and 4 show rate of hydrolysis data of various inventive compounds in human intestinal homogenates.

Figures 5 and 6 are plots of the rate of hydrolysis of inventive compound in human liver homogenates.

25 Figure 7 is a plot of the concentrations of lisofylline and its corresponding S enantiomer in canine blood over time, after oral administration of inventive compound no. 4544R.

Figure 8 is a plot of the concentrations of lisofylline and its corresponding S enantiomer in canine blood over time, after oral administration of inventive compound no. 4545R.

30 Figure 9 is a plot of the concentrations of lisofylline and its corresponding S enantiomer in canine blood over time, after oral administration of inventive compound no. 4576R.

Figure 10 is a plot of concentration of lisofylline and its corresponding S enantiomer over 2 hours in canine blood dosed with inventive compound no. 4577R.

Detailed Description of Preferred Embodiments

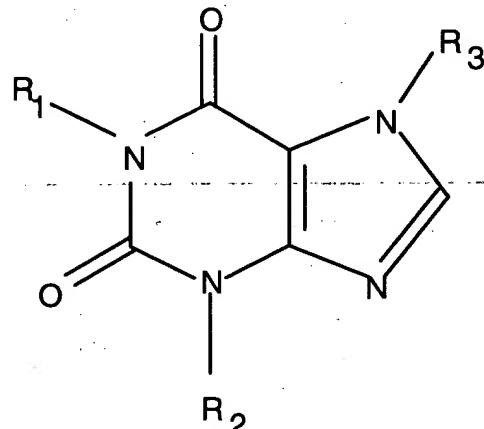
35 The inventive compounds include, *inter alia*, compounds that form the hydroxyl-substituted xanthine, lisofylline *in vivo*. Of the various advantages of the compounds, these

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compounds have a primary, characteristic benefit: selective enantiomeric stability coupled with varying resistance to hydrolysis. Having performed detailed studies of lisofylline, its *in vivo*, metabolic pathway and corresponding affect on its therapeutic potential, a class of novel compounds was discovered having structures and chemical properties that were previously unknown.

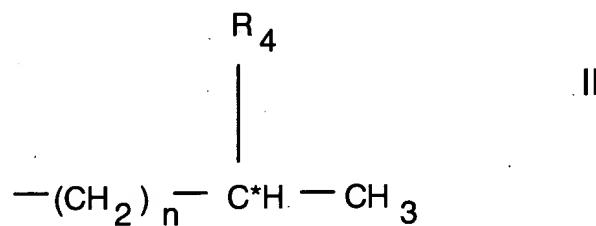
Compounds and Compositions

The present invention provides a chiral ω -1 hydroxyl substituted xanthine compound comprising a xanthine core of formula I:



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One of R_1 or R_2 is independently an aliphatic hydrocarbon having the formula II:



15 In formula II, n is an integer from about four to about eight, R_1 or R_2 , which is other than formula II, and R_3 are independently $C_{(1-12)}$ alkyl and C^* is a chiral carbon atom. R_4 is an amino acid or carbohydrate attached to the chiral carbon atom C^* via an ester linkage, or
 $-O-X-(R_5)_m$.
If R_4 is $-O-X-(R_5)_m$, m is two or three, X is selected from the group consisting of C , P
20 or S and R_5 is a hydrogen atom; hydroxyl group; $=O$; substituted or unsubstituted $C_{(1-10)}$ alkyl, $C_{(1-10)}$ alkenyl, $C_{(1-10)}$ alkynyl, $C_{(1-10)}$ alkoxyl, $C_{(1-10)}$ oxoalkyl, or $C_{(1-10)}$ acetoxy, $C_{(1-10)}$ carboxyalkyl, or $C_{(1-10)}$ hydroxyalkyl group; $-OR_6$, R_6 being a substituted or unsubstituted $C_{(1-10)}$ alkyl, $C_{(1-10)}$ alkenyl, $C_{(1-10)}$ alkynyl; $C_{(1-10)}$ oxoalkyl; substituted

or unsubstituted cyclic or heterocyclic group having from one- to three-rings, each ring containing from four to seven atoms.

Representative amino acids may be naturally occurring or synthesized. Examples of amino acids include, without limitation, alaninyl, argininy, asparaginyl, aspartyl, cysteinyl, glutaminyl, glutamyl, glyciny, histidinyl, isoleucinyl, leucinyl, lysinyl, methioninyl, phenylalaninyl, prolinyl, serinyl, threoninyl, tryptophanyl, tyrosinyl, valinyl. Representative, non-limiting carbohydrates include glucosyl, glucosidyl, maltosyl, glucopyranosidyl, glyceraldehydyl, erythrosyl, arabinosyl, ribulicosyl, fructosyl, erythritolyl, xylosyl, lyxosyl, allosyl, altrosyl, mannosyl, mannosidyl, gulosyl, idosyl, galactosyl or talosyl.

If R₄ is -O-X-(R₅)_m, m is two or three, X is selected from the group consisting of C, P or S and R₅ is a hydrogen atom; hydroxyl group; =O; substituted or unsubstituted C₍₁₋₁₀₎ alkyl, C₍₁₋₁₀₎ alkenyl, C₍₁₋₁₀₎ alkynyl, C₍₁₋₁₀₎ alkoxy, C₍₁₋₁₀₎ oxoalkyl, or C₍₁₋₁₀₎ acetoxy or C₍₁₋₁₀₎ hydroxyalkyl group; -OR₆, R₆ being a substituted or unsubstituted C₍₁₋₁₀₎ alkyl, C₍₁₋₁₀₎ alkenyl, C₍₁₋₁₀₎ alkynyl or C₍₁₋₁₀₎ oxoalkyl; substituted or unsubstituted cyclic or heterocyclic group having from one- to three-rings, each ring containing from four to seven atoms.

In preferred inventive compounds, X is C. If R₄ is -O-X-(R₅)_m, more preferred compounds have at least one R₅ that is =O and m is two.

Representative, non-limiting examples of substituents for the substituted C₍₁₋₁₀₎ alkyl, C₍₁₋₁₀₎ alkenyl, C₍₁₋₁₀₎ alkynyl, C₍₁₋₁₀₎ alkoxy, C₍₁₋₁₀₎ oxoalkyl, or C₍₁₋₁₀₎ acetoxy, cyclic or heterocyclic groups include: amido, amino, C₍₁₋₆₎ alkenyl, C₍₁₋₆₎ alkyl, C₍₁₋₆₎ alkoxy, primary, secondary or tertiary C₍₁₋₆₎ hydroxyalkyl, C₍₁₋₆₎ oxoalkyl, azido, carbonyl, carboxylic acid, cyano, C₍₁₋₆₎ haloalkyl, isocyano, isothiocyanato, phosphatyl, phosphonatyl, sulfonatyl, sulfonyl, sulfoxyl, imino, thioamido, thiocarbonyl, thioalkoxyl, thioloalkyl and thio groups or a single atom. The single atom may be oxygen, chlorine, bromine or fluorine. The C₍₁₋₆₎ haloalkyl may be a mono-, di- or tri-haloalkyl and the C₍₁₋₆₎ alkoxy may preferably be a methoxy or ethoxy group.

Most preferred compounds of the invention have the R₁ or R₂, other than formula II, containing one or two, nonadjacent oxygen atoms, each oxygen atom replacing a single carbon atom of the C₍₁₋₁₂₎ alkyl.

Preferred cyclic or heterocyclic groups of the inventive compounds may include, but are not limited to, a benzyl, phenyl, biphenyl, cyclohexyl, cyclohexenyl, cyclopentyl, cyclopentenyl, cyclopantanediyl, napthalenyl, phenyl, quinonyl, cyclopropyl, cyclobutyl, cycloheptyl, cycloheptenyl, indanyl, indenyl, decalinyl, resorcinolyl, tetralinyl, α-tetralonyl, 1-indanonyl, cyclohexanediyl, cyclopantanediyl, dimethylxanthinyl, methylxanthinyl, phthalimidyl, homophthalimidyl, methylbenzoyleneureayl, quinazolinonyl,

octylcarboxamidobenzeryl, methylbenzamidyl, methyldioxotetrahydropteridinyl, glutarimidyl, piperidonyl, succinimidyl, dimethoxybenzetyl, methyldihydouracilyl, methyluracilyl, methylthymaryl, piperidinyl, dihydroxybenzetyl or methylpurinyl, even more preferably, methylxanthinyl or dimethylxanthinyl group.

5 In most preferred compounds of the invention, n is 4, m is 2, and R₂ and R₃ are methyl and at least one R₅ is =O. The other R₅, other than =O, preferably may be selected from the group consisting of trimethoxy-substituted phenyl, phenolyl or benzamino. Most preferred of the amino acid, R₄, have R₄ selected from among glycanyl, isoleucanyl and valanyl.

10 Pharmaceutical Formulations

A suitable pharmaceutical formulation will depend on the nature of the disorder to be treated, the nature of the medicament chosen, and the judgment of the attending physician. In general, the compounds are formulated for oral administration. Suitable formulations for these compounds can be found, for example, in *Remington's Pharmaceutical Sciences* (latest edition), Mack Publishing Company, Easton, PA.

15 The compounds, their pharmaceutically acceptable salts, solvates and hydrates can be employed in a wide variety of pharmaceutical forms. The preparation of a pharmaceutically acceptable salt will be determined by the chemical nature of the compound itself, and can be prepared by conventional techniques readily available. Thus, if a solid carrier is used, the preparation can be tableted, placed in a hard gelatin capsule in powder or pellet form or in the form of a troche or lozenge. The amount of solid carrier will vary widely but preferably will be from about 25 mg to about 1 gram, wherein the amount of inventive compound per dose will vary from about 25 mg to about 1 gram for an adult. When a liquid carrier is used, the preparation will be in the form of a syrup, emulsion, soft gelatin capsule, sterile injectable liquid such as an ampule or nonaqueous liquid suspension. Where the inventive composition is in the form of a capsule, any routine encapsulation is suitable, for example, using the aforementioned carriers in a hard gelatin capsule shell. Where the composition is in the form of a soft gelatin shell capsule, any pharmaceutical carrier routinely used for preparing dispersions of suspensions may be considered, for example, aqueous gums, celluloses, silicates or oils and are incorporated in a soft gelatin capsule shell. A syrup formulation will generally consist of a suspension or solution of the compound or salt thereof in a liquid carrier (e.g., ethanol, polyethylene glycol, coconut oil, glycerine or water) with a flavor or coloring agent.

20 The amount of inventive compound required for therapeutic effect upon administration will, of course, vary with the compound chosen, the nature and severity of the disease and the discretion of the treatment provider. The daily dosage for treatment of sepsis or another severe inflammatory condition by oral administration from about 0.001 mg/kg to about 40

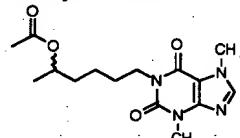
mg/kg, preferably from about 0.01 mg/kg to about 20 mg/kg of an inventive compound or a pharmaceutically acceptable salt thereof, calculated as the free base.

The daily dosage regimen for oral administration is suitably from about 0.1 mg/kg to about 1000 mg/kg per day. For administration the dosage is suitably from about 0.001 mg/kg to about 40 mg/kg of the inventive compound or a pharmaceutically acceptable salt thereof, calculated as the free base. The active ingredient may be administered from 1 to 6 times a day, sufficient to exhibit activity.

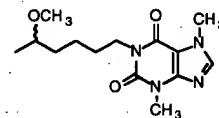
While dosage values will vary, therapeutic compounds of the invention may be administered to a human subject requiring such treatment as an effective oral dose of about 50 mg to about 5000 mg per day, depending upon the weight of the patient. For any particular subject, specific dosage regimens should be adjusted to the individual's need and to the professional judgment of the person administering or supervising the administration of the inventive compounds.

Representative compounds according to the invention may include:

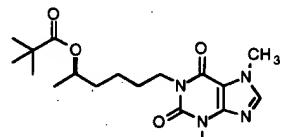
15 1515R 1-(5-Acetoxyhexyl)-3,7-dimethylxanthine



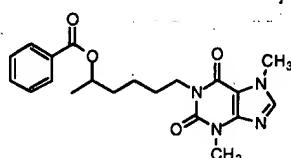
1570R 1-(5-(R)-methoxyhexyl)-3,7-dimethylxanthine



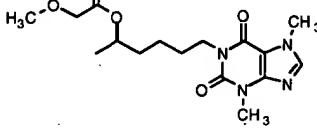
4544R R-1-(5-Pivaloyloxyhexyl)-3,7-dimethylxanthine



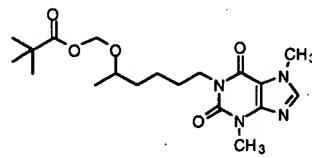
4545R R-1-(5-Benzoyloxyhexyl)-3,7-dimethylxanthine

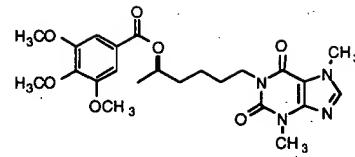


4546R R-1-(5-Methoxyacetoxyhexyl)-3,7-dimethylxanthine

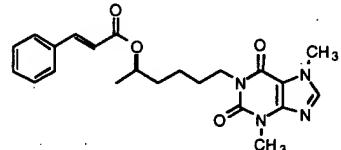


20 4547R R-1-(5-(Pivaloyloxymethoxyhexyl)-3,7-dimethylxanthine

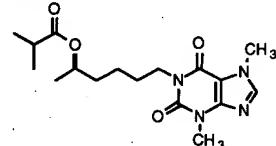




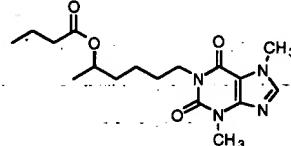
4570R R-1-(5-(3,4,5-Trimethoxybenzoyloxy)hexyl)-3,7-dimethylxanthine



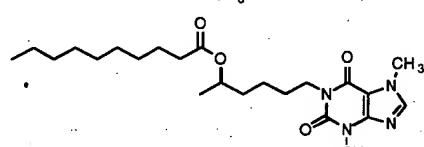
4571R R-1-(5-trans-Cinnamoyloxy)hexyl)-3,7-dimethylxanthine



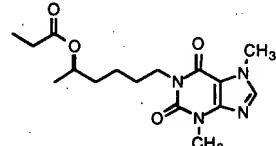
4572R R-1-(5-(Isobutyroxy)hexyl)-3,7-dimethylxanthine



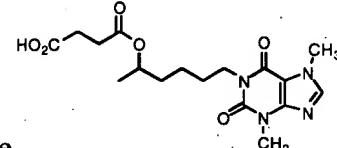
5 4573R R-1-(5-Butyroxy)hexyl)-3,7-dimethylxanthine



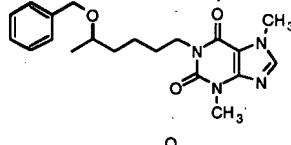
4574R R-1-(5-Decanoxy)hexyl)-3,7-dimethylxanthine



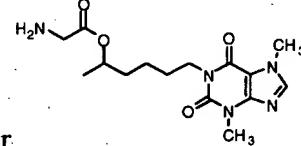
4575R R-1-(5-Propionoxy)hexyl)-3,7-dimethylxanthine



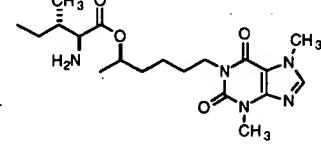
4576R R-1-(5-(3-Carboxy)propionoxy)hexyl)-3,7-dimethylxanthine



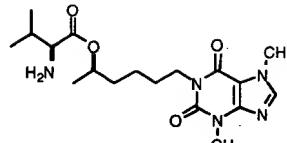
4577R R-1-(5-Benzyl)hexyl)-3,7-dimethylxanthine



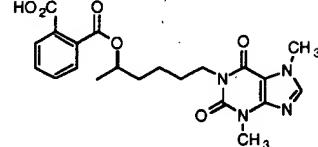
10 5500R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine glycinate ester



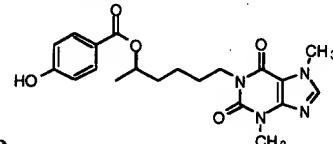
5501R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine isoleucinate ester



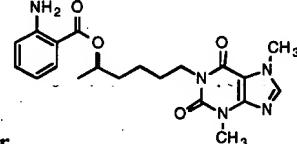
5502R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine valinate ester



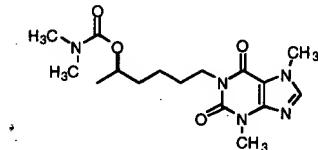
5503R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine phthalic acid half ester



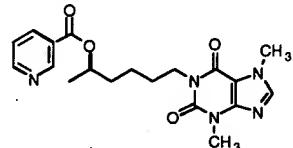
5504R R-1-(5-(4'-Hydroxybenzoyloxy)hexyl)-3,7-dimethylxanthine



5 5527R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine anthranilate ester



5531R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine N,N-dimethyl carbamate



5540R R-1-(5-Hydroxy)hexyl)-3,7-dimethylxanthine nicotinate ester

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Although various synthetic protocols may be used by skilled artisans to prepare the inventive compounds, generally, the compounds may be prepared by reacting a starting material, lisofylline, with a compound having a structure of the functional moiety (R₄ of the above formula II) of these compounds. Thus, providing that starting materials are commercially available or may be readily synthesized, a variety of compounds within the scope of the invention may be synthesized by using this general method, selecting starting materials and reagents to achieve a desired product. Figure 1 graphically represents this general synthetic process.

The following examples are illustrative of specific, preferred embodiments of the invention; however, these examples are not intended to be construed as limiting the scope of the invention as disclosed herein.

Example 1

This example illustrates several methods of synthesizing inventive compound no. 1515R--protocols A-D, below--(see above for chemical name and structure).

A. Inventive compound no. 1515 may be prepared from lisofylline via acid catalysis. A 200 mL round bottom ("r.b.") flask was charged with lisofylline (28 g, 0.1 mol; 99% chemical purity, 93% R isomer) and acetic anhydride (51 g, 0.5 mol). The mixture was magnetically stirred and concentrated sulfuric acid (0.5 mL) was added dropwise to the stirring solution. The resulting reaction mixture was heated to reflux and maintained at reflux temperature for 16 hours. The hot mixture was drowned in ice (200 g), and the mixture neutralized and saturated with sodium bicarbonate (80 g), yielding a slurry containing a tan solid. The tan solid was taken up in ethyl acetate (500 mL) and the aqueous layer was separated and discarded.

The organic layer was washed with two, 250 mL aliquots of saturated brine, and the brine layers discarded. The organic layer was dried over sodium sulfate, filtered through a small plug of neutral alumina to remove colored impurities, and stripped to a yellow/orange thick syrup, which crystallizes on standing to an off-white solid, leaving 35.6 g of crude yield. The crude product was recrystallized from 1.5 parts ethyl acetate and 3 parts hexanes, yielding 15.4 g of colorless to off-white fluffy needles--1515R--(47.8%, 99.2% chemical purity by HPLC).

B. Inventive compound no. 1515 was also prepared from lisofylline via base catalysis. A 500 mL r.b. flask was charged with lisofylline (50 g, 0.178 mol), ethyl acetate (100 mL), triethylamine (21.6 g, 0.214 mol) and 4-dimethylaminopyridine (0.217 g, 1 mol%). The mixture was slurried and chilled to 5 °C in an ice water bath, followed by slow addition of acetic anhydride (20 g, 0.196 mol) to the mixture over 10 minutes. The slurry was allowed to warm to ambient temperature over 1 hour and was stirred at room temperature for 16 hours. The slurry became homogeneous, and TLC indicated a single spot for product. The mixture was drowned with water (150 mL) and additional ethyl acetate (100 mL). The mixture was separated and the aqueous layer was removed and discarded. The organic layer was washed with saturated brine (100 mL) and the brine layer discarded. The organic layer was dried over sodium sulfate and stripped to a yellow residue, which crystallized upon standing to yield 60.2 g of a crude off-white solid. The crude product was recrystallized from ethyl acetate/petroleum ether (30-60 °C boiling range), resulting in 29 g of colorless fluffy needles (51% yield, m.p.=81.5-83.5, 99.8% chemical purity by HPLC).

C. Inventive compound no. 1515 was also prepared from 1-chloro-5-(R)-acetoxyhexane according to the following synthetic method. Under a slow sweep of dry nitrogen, a 250 mL, 3-neck, round-bottomed flask fitted with a reflux condenser, thermometer

and pressure equalizing addition funnel was charged with sodiotheobromine (10.2 g, 50.5 mmol), and 30 mL DMSO (Gaylord high tech grade, 99.9% minimum.). The resulting slurry was stirred at ambient temperature and the vessel was charged with 1-chloro-5-(R)-acetoxyhexane (9.66g, 50.5 mmol; 93% chemical purity). No exothermic reaction was
5 observed.

The resulting mixture was heated to 80 °C and held at that temperature for a minimum of 6 hours. The reaction mixture cleared up over the 6 hour period to a hazy yellow solution. The mixture was drowned with water (30 mL) and cooled to ambient temperature. The mixture was drowned with isopropyl acetate (60 mL) and the mixture was stirred for 10
10 minutes to extract the desired product, after which the mixture was separated and the lower aqueous DMSO layer was discarded. The organic layer was washed with saturated brine (30 mL), the mixture was separated, and the brine layer discarded. The solvent of the organic layer was distilled off, leaving a thick syrup, which crystallized upon standing to obtain 14.5 g
15 of crude product. The crude product was recrystallized from isopropyl acetate/ isoctane in 76% yield and 96.5% chemical purity.

D: Inventive compound no. 1515 was also prepared from 1-bromo-5-(R)-acetoxyhexane according to the following synthetic method. The synthetic procedure described in detail in paragraph C above was repeated, except that the resulting slurry (after addition of sodiotheobromine) was charged with 1-bromo-5-(R)-acetoxyhexane, instead of 1-chloro-5-(R)-acetoxyhexane. In addition, the reaction mixture was heated at 55-60 °C for 16-24 hours, and the reaction mixture was drowned with up to 60 mL water. The desired product was instead extracted with toluene or ethyl acetate instead of isopropyl acetate.
20

Example 2

This method illustrates a method of synthesis for inventive compound no. 1570R (see above for chemical names and structure). A 250 mL two-necked flask fitted with an argon inlet and a septum was charged with (95%) sodium hydride (360 mg; 15 mmol). A solution of lisofylline (2.8 g, 10 mmol) in tetrahydrofuran (40 mL) was added dropwise and the resulting reaction mixture stirred for 30 minutes. Methyl iodide (2.838 g; 20 mmol) was added and the reaction mixture stirred for 12 hours. The solvent was then removed under reduced pressure and water (50 mL) was added. The reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with brine solution (50 mL) dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.8 g of compound no. 1570R (yield = 61%).
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30
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Example 3

This example illustrates a method of synthesis for inventive compound no. 4544R (see above for chemical name and structure). A 250 mL r.b. flask fitted with a dropping funnel and a drying tube was charged with a solution of lisofylline (2.8 g, 10 mmol) in tetrahydrofuran (40 mL), triethylamine (10 mL) and N,N-dimethylaminopyridine (100 mg). The reaction flask was cooled to 10 °C and pivolyl chloride (3 mL, 24 mmol) was added dropwise and stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL), brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant, producing 2.4 g of inventive compound no. 4544R (yield = 67%).

Example 4

This example illustrates a method of synthesis for inventive compound no. 4545R (see above for chemical name and structure). A 250 mL r.b. flask fitted with a dropping funnel and a drying tube was charged with a solution of lisofylline (2.8 g, 10 mmol) in tetrahydrofuran (40 mL), triethylamine (10 mL) and N,N-dimethylaminopyridine (100 mg). The reaction flask was cooled to 10 °C and benzoyl chloride (2.1 g; 1.74 mL, 15 mmol) was added dropwise. The resulting mixture was stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL), cold dilute hydrochloric acid (50 mL) and brine solution (50 mL); dried over anhydrous magnesium sulfate; and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant, producing 3.0 g of inventive compound no. 4545R (yield = 78%).

Example 5

This example illustrates a method of synthesis for inventive compound no. 4546R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (2.8 g, 10 mmol) in dichloromethane (40 mL) and triethylamine (10 mL). The reaction flask was cooled to 10 °C and methoxyacetyl chloride (1.63 g; 1.4 mL; 15 mmol) was added dropwise and stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under

reduced pressure. The crude product obtained was then further purified by flash chromatography over silica gel using hexane(20%)/ethyl acetate as eluant, resulting in 2.0 g of compound no. 4546R (yield = 57%).

5 **Example 6**

This example illustrates a method of synthesis for inventive compound no. 4547R (see above for chemical name and structure). A 250 mL two-necked flask fitted with an argon inlet and a septum was charged with (95%) sodium hydride (360 mg; 15 mmol). A solution of lisofylline (2.8 g, 10 mmol) in tetrahydrofuran (40 mL) was added dropwise and stirred for 30 minutes. Chloromethylpivalate (2.26 g; 15 mmol) was added and a resulting reaction mixture was refluxed for 8 hours. The solvent was then removed under reduced pressure. Water (50 mL) was slowly added at room temperature and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant, resulting in 2.9 g of inventive compound no. 4547R (yield = 74%).

Example 7

20 This example illustrates a method of synthesis for inventive compound no. 4570R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL) and triethylamine (5 mL). The reaction flask was cooled to 10 °C and 3,4,5-trimethoxybenzoyl chloride (1.15 g; 5mmol) was added dropwise. The resulting reaction mixture was stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL), cold dilute hydrochloric acid (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate/20% hexane as eluant to obtain 2.0 g of compound no. 4570R (yield = 57%).

Example 8

35 This example illustrates a method of synthesis for inventive compound no. 4571R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL) and triethylamine (5 mL). The reaction flask was cooled to 10 °C and trans-

cinnamoyl chloride (1.0 g; 6 mmol) was added in portions and stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure.

5 The crude product obtained was further purified by flash chromatography over silica gel using an hexane(10%)/ethyl acetate as eluant, resulting in the 1.6 g of compound no. 4571R (yield = 78%).

Example 9

10 This example illustrates a method of synthesis for inventive compound no. 4572R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL), triethylamine (5 mL) and NN-dimethylaminopyridine (100 mg). The reaction flask was cooled to 10 °C and isobutyric anhydride (1.58 g; 1.65 mL; 10 mmol) was added dropwise, the resulting mixture stirred for 24 hours. The reaction was quenched by the addition of methanol (5 mL), followed by water (50 mL). The reaction mixture was extracted with four 50 mL aliquots of ethyl acetate and the combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by

15 flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.6 g of compound no. 4572R (yield = 91%).

Example 10

20 This example illustrates a method of synthesis for inventive compound no. 4573R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL), triethylamine (5 mL) and N,N-dimethylaminopyridine (100 mg). The reaction flask was cooled to 10 °C and butyric anhydride (1.58 g; 1.65 mL; 10 mmol) was added dropwise and stirred for 24 hours. The reaction was quenched by the addition of methanol (5 mL)

25 followed by water (50 mL). The reaction mixture was extracted with four 50 mL aliquouts of ethyl acetate. The combined organic extracts were subsequently washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant, resulting in 1.48 g of compound no. 4573R (yield =

30 87%).

Example 11

This example illustrates a method of synthesis for inventive compound no. 4574R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL) and triethylamine (5 mL). The reaction flask was cooled to 10 °C and decanoyl chloride (1.14 g; 6 mmol) was added in portions. The resulting mixture was stirred for 24 hours. Water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using an hexane(20%)/ethyl acetate as eluant to obtain 1.6 g of compound no. 4574R (yield = 74%).

Example 12

This example illustrates a method of synthesis for inventive compound no. 4575R (see above for chemical name and structure). A 250 mL r.b. flask fitted with a dropping funnel and a drying tube was charged with a solution of lisofylline (1.4 g, 5 mmol) in dichloromethane (20 mL), triethylamine (5 mL) and N,N-dimethylaminopyridine (100 mg). The reaction flask was cooled to 10 °C and propionic anhydride (0.79 g; 6 mmol) was added dropwise and stirred for 24 hours. The reaction was quenched by the addition of methanol (5 mL), followed by water (50 mL). The reaction mixture was extracted with four 50 mL aliquots of ethyl acetate and the combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.2 g of inventive compound no. 4575R (yield = 71%).

Example 13

This example illustrates a method of synthesis for inventive compound no. 4576R (see above for chemical name and structure). A 250 mL r.b. flask fitted with a dropping funnel and a drying tube was charged with a solution of lisofylline (1.40 g, 5mmol) in ethyl acetate (20 mL), triethylamine (1.4 mL;10 mmol) and N,N-dimethylaminopyridine (122 mg; 1 mmol). The reaction flask was cooled to 10° C and succinic anhydride (0.6 g, 6 mmol) was added. The resulting mixture was stirred for 24 hours. 2N hydrochloric acid (10. mL) was added and the reaction mixture was extracted with four 50 mL aliquots of dichloromethane. The combined organic extracts were washed with brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further

purified by flash chromatography over silica gel using a 5% methanol/ethyl acetate eluant, producing 1.0 g of inventive compound no. 4576R (yield = 53%).

Example 14

5 This example illustrates a method of synthesis for inventive compound no. 4577R (see above for chemical name and structure). A 250 mL two-necked flask, fitted with an argon inlet and a septum, was charged with (95%) sodium hydride (180 mg; 7.5 mmol). A solution of lisofylline (1.4 g, 5 mmol) in tetrahydrofuran (20 mL) was added dropwise and the reaction mixture stirred for thirty minutes. Benzyl bromide (1.28 g; 0.895 mL; 7.5 mmol) was added
10 and the reaction mixture was refluxed for 8 hours. The solvent was then removed under reduced pressure and water (50 mL) was added at room temperature slowly. The reaction mixture was extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by
15 flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.0 g of compound no. 4577R (yield = 1.0 g (54 %)).

Example 15

20 This example illustrates a method of synthesis for inventive compound no. 5500R (see above for chemical name and structure). A 100 mL two-necked flask, fitted with an argon inlet and a septum, was charged with a solution of lisofylline (1.4 g, 5 mmol) N-carbobenzyloxyglycine (1.225 g; 6 mmol) and N,N-dimethylaminopyridine (488.7 mg; 4 mmol) in dry dichloromethane (20 mL). A 1M solution of dicyclohexylcarbodiimide (6 mL; 6mmol) in dichloromethane was added and stirred for 16 hours. The precipitated urea was removed by
25 filtration and the residue was washed with dichloromethane (50 mL). The combined filtrate was concentrated under reduced pressure and the crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.8 g of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl glycinate ester (yield = 76%).

30 A solution of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl glycinate ester (1.8 g; 3.82 mmol), prepared above, in ethanol (50 mL) was hydrogenated in presence of 10% palladium on carbon (400 mg) at room temperature for 12 hours. The reaction mixture was then filtered and concentrated under reduced pressure. The crude product was then redissolved in dichloromethane (15 mL) and treated with 1M solution of hydrogen chloride in ether (5 mL). The solvent was removed under reduced pressure to obtain
35 1.05 g of compound no. 5500R (yield = 74%).

Example 16

This example illustrates a method of synthesis for inventive compound no. 5501R (see above for chemical name and structure). A 100 mL two-necked flask, fitted with an argon inlet and a septum, was charged with a solution of lisofylline (1.4 g, 5 mmol) N-carbobenzyloxy-L-isoleucine (1.6 g; 6 mmol) and N,N-dimethylaminopyridine (488.7 mg; 4 mmol) in dry dichloromethane (20 mL). A 1M solution of dicyclohexylcarbodiimide (6 mL; 6 mmol) in dichloromethane was added and stirred for 16 hours. The precipitated urea was removed by filtration and the residue was washed with dichloromethane (50 mL). The combined filtrate was concentrated under reduced pressure and the crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant to obtain 2.4 g of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl-L-isoleucinate ester (yield = 91%).

A solution of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl-L-isoleucinate ester (2.24 g; 4.25 mmol), prepared above in ethanol (50 mL) was hydrogenated in the presence of 10% palladium on carbon (400 mg) at room temperature for 12 hours. The reaction mixture was then filtered and concentrated under reduced pressure. The crude product was then redissolved in dichloromethane (15 mL) and treated with 1M solution of hydrogen chloride in ether (5 mL). The solvent was removed under reduced pressure to obtain 1.2 g of compound no. 5501R (yield = 71%).

20

Example 17

This example illustrates a method of synthesis for inventive compound no. 5502R (see above for chemical name and structure). A 100 mL two-necked flask, fitted with an argon inlet and a septum, was charged with a solution of lisofylline (1.4 g, 5 mmol) N-carbobenzyloxy-L-valine (1.5 g; 6 mmol) and N,N-dimethylaminopyridine (488.7 mg; 4 mmol) in dry dichloromethane (20 mL). A 1M solution of dicyclohexylcarbodiimide (6 mL; 6 mmol) in dichloromethane was added and stirred for 16 hours. The precipitated urea was removed by filtration and the residue was washed with dichloromethane (50 mL). The combined filtrate was concentrated under reduced pressure and the crude product obtained was further purified by flash chromatography over silica gel using ethyl acetate as eluant to obtain 1.9 g of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl-L-valinate ester (yield = 1.9 g 76%).

A solution of 1-(5-(R)-hydroxyhexyl)-3,7-dimethylxanthine N-benzyloxycarbamoyl-L-valinate ester (1.8 g; 3.5 mmol) in ethanol (50 mL) was hydrogenated in presence of 10% palladium on carbon (400 mg) at room temperature for 12 hours. The reaction mixture was then filtered and concentrated under reduced pressure. The crude product was then

redissolved in dichloromethane (15 mL) and treated with 1M solution of hydrogen chloride in ether (4 mL). The solvent was removed under reduced pressure, resulting in 1.2 g of inventive compound no. 5502R (yield = 83%).

5 **Example 18**

This example illustrates a method of synthesis for inventive compound no. 5503R (see above for chemical name and structure). A 250 mL r.b. flask fitted with a dropping funnel and a drying tube was charged with a solution of lisofylline (1.40 g, 5mmol) in dichloromethane (30 mL), triethylamine (1.4 mL, 10 mmol) and NN-dimethylaminopyridine (122 mg; 1 mmol).

10 The reaction flask was cooled to 10 °C, after which phthalic anhydride (0.89 g, 6 mmol). The resulting reaction mixture was stirred for 24 hours. 2N hydrochloric acid (10 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of dichloromethane. The combined organic extracts were washed with brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. A crude product from this 15 concentration was further purified by flash chromatography over silica gel using a 5% methanol/ethyl acetate eluant. Recrystallization of the purified product from ethyl acetate produced 1.4 g of compound 5503 (yield = 65%).

Example 19

20 This example illustrates a method of synthesis for inventive compound no. 5504R (see above for chemical name and structure). 4-Benzylxybenzoic acid was first prepared (paragraph A below) and then used to make compound 5504 (paragraph B below).

A) 4-benzylxybenzoic acid:

25 A solution of 4-hydroxybenzoic acid (5.52 g; .40 mole), benzyl bromide (11.89 mL; 0.1 mol) and tricaprylylmethylammonium chloride (100 mg, Aliquat 336[®] Henkel Corporation) in methanol (50 mL) was refluxed in presence of potassium carbonate (16.52 g) for 12 hours. The reaction mixture was then poured slowly into a solution of 2N hydrochloric acid solution. The precipitated acid was filtered, dried and recrystallized from hexane/ethyl acetate to give 5 g of 4-benzylxybenzoic acid (yield = 55%).

30 B) 1-(5-(R)-(4'-benzylxybenzyloxy)hexyl)-3,7-dimethylxanthine:

A 100 mL two-necked flask, fitted with an argon inlet and a septum, was charged with a solution of lisofylline (2.8 g, 10 mmol), 4-benzylxybenzoic acid, prepared above, (2.73 g; 1.2 mmol) and N,N-dimethylaminopyridine (977 mg; 8 mmol) in dry dichloromethane (30 mL). A 1M solution of dicyclohexylcarbodiimide (12 mL; 12 mmol) in dichloromethane was added 35 and the resulting reaction mixture stirred for 16 hours. The precipitated urea was removed by

filtration and the residue was washed with dichloromethane (50 mL). The combined filtrate was concentrated under reduced pressure and the crude product obtained was further purified by flash chromatography over silica gel using an ethyl acetate/30% hexane eluant, resulting in 4.45 g 1-(5-(R)-(4'-benzyloxybenzoyloxy)hexyl)-3,7-dimethylxanthine (yield = 91%).

5 A solution of 1-(5-(R)-(4'-benzyloxybenzoyloxy)hexyl)-3,7-dimethylxanthine (4.48 g) in ethanol (50 mL) was hydrogenated in presence of 10% palladium on carbon (800 mg) at room temperature for 12 hours. The reaction mixture was then filtered and concentrated under reduced pressure to obtain 3.2 g of compound no. 5504R (yield = 3.2 g 88%).

10 **Example 20**

This example illustrates a method of synthesis for inventive compound no. 5527R (see above for chemical name and structure). A 250 mL r.b. flask, fitted with a dropping funnel and a drying tube, was charged with a mixture of lisofylline (2.8 g, 10 mmol), isatoic anhydride (2 g, 12 mmol), NN-dimethylaminopyridine (1.2 g 10 mmol) and dimethylformamide (25 mL).

15 The reaction flask was heated to 80 °C for 4 hours. After cooling to room temperature, water (50 mL) was added and the reaction mixture was extracted with four 50 mL aliquots of ethyl acetate/10% methanol. The combined organic extracts were washed with water (50 mL) and brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography 20 over silica gel using ethyl acetate as eluant, resulting in 0.72 g of compound no. 5527R (yield = 18 %).

Example 21

This example illustrates a method of synthesis for inventive compound no. 5531R (see above for chemical name and structure). A 250 mL two-necked flask, fitted with an argon inlet and a septum, was charged with (95%) sodium hydride (144 mg; 6 mmol). A solution of lisofylline (1.4 g, 5 mmol) in tetrahydrofuran (20 mL) was added dropwise and stirred for 30 minutes. N,N-dimethylcarbomyl chloride (0.645 g; 6 mmol) was added and the reaction mixture was refluxed for 12 hours. The reaction mixture was then cooled back to room 30 temperature, and water (50 mL) was added slowly. The reaction mixture was then extracted with four 50 mL aliquots of ethyl acetate. The combined organic extracts were washed with brine solution (50 mL), dried over anhydrous magnesium sulfate and concentrated under reduced pressure. The crude product obtained was further purified by flash chromatography over silica gel using 50% dichloromethane/49% hexane/1% methanol as eluant, which resulted 35 in 1.3 g of compound no. 5531R (yield = 74 %).

Example 22

This example provides data on *in vitro* metabolism of lisofylline, its corresponding opposite enantiomer, 1-[5-(S)-hydroxyhexyl]-3,7-dimethylxanthine, and stability of the inventive compounds nos. 1515R, 1570R, 4544R, 4545R, 4546R and 4547R, when incubated 5 with samples of human whole blood.

A stereoselective HPLC method was used to determine concentrations of lisofylline and its corresponding opposite isomer. Separation of analytes as their phenylisocyanate derivatives was accomplished by HPLC on a Chiracel OJ (25 x 0.46 cm, 5 μ) analytical column with UV detection at 273 nm. The mobile phase was methanol/water/trifluoroacetic acid 10 (95:4.5:0.5) delivered at a flow rate of 0.8 mL/minute with a run time of 30 minutes. The column temperature was set at 30 °C. Data was collected using the Waters 860 Chromatography Data System with Expert Ease 3.2 software. Calibration was accomplished by a weighted 1/x least squares regression analysis. Calculations were based on peak area ratios. Total recovery of analytes [the combined R and S enantiomers of corresponding 15 inventive compound and lisofylline and its corresponding S enantiomer] in these experiments was 80%-85%.

Procedurally, whole human blood was obtained from healthy volunteers and collected into pre-wetted tubes with 3.8% sodium citrate as anticoagulant. The final concentration of inventive compounds in the incubation was 100 μ M. Incubation time was 60 minutes for all 20 tissues (bath temperature: 37 °C; shaker setting: 30 rpm). Samples were taken at the following time intervals: 0, 5, 15, 30 and 60 minutes. Enzymatic activity was arrested by pre-wetting the sample culture tubes with 50 mL of acetone, and cooling the sample tubes in ice.

Figure 2 reports results obtained in this assay, illustrating amounts of hydrolysis of various inventive compounds with whole human blood. The extent of hydrolysis is measured 25 by the appearance of lisofylline and was measured by the stereoselective HPLC method described above. As shown by the data reported in figure 2, compounds 1570R, 4544R and 4545R are quite resistant to hydrolysis in incubations with whole human blood. In comparison, compounds nos. 1515R and 4546R are hydrolyzed quite readily under these conditions and compound no. 4547R has an hydrolysis rate between the most resistant 30 compounds tested and those readily hydrolyzed, indicating that a specific rate of hydrolysis can be selected for a particular use of the compound.

Example 23

This example provides data on the *in vitro* hydrolysis of representative inventive 35 compounds in human intestinal homogenates.

Procedurally, homogenate was prepared in DMEM : F-12 Hams (50: 50) with essential trace elements (commercial preparation), NaHCO₃ and HEPES (pH 7.0). Tissue was weighed to the nearest 0.1 mg and minced on an iced petri dish. Minced tissue was placed in an iced homogenizer (with 10 mL media) and disrupted with ten (10) strokes of a motorized Teflon 5 pestle. The volume of homogenate was adjusted with the appropriate amount of media for each incubation set. Approximate protein concentration was 30 mg/mL. Final incubation volume per compound was 3 mL. Compound concentrations were identical to compound concentration used in Example 22. Lisofylline was incubated under the same conditions and concentration used for the inventive compounds tested to demonstrate recovery of lisofylline in 10 the system.

The extent of hydrolysis was measured by the appearance of lisofylline and is measured by the stereoselective HPLC method described in Example 22, above.

Figures 3 and 4 illustrate the hydrolysis of representative compounds in the presence of human intestinal homogenates. The compound most resistant to hydrolysis was 1570R, followed by 4544R and 4545R. Compound no. 4547R was hydrolyzed at an intermediate rate while 1515R and 4546R underwent rapid hydrolysis in comparison to other compounds tested. This trend is similar to that observed in whole human blood.

Example 24

This example provides data on the *in vitro* hydrolysis of representative inventive compounds in human liver homogenates.

Homogenate was prepared in DMEM : F-12 Hams (50: 50) with essential trace elements (commercial preparation), 14.3 mM NaHCO₃ and 15 mM HEPES (pH 7.0). Tissue was weighed to the nearest 0.1 mg and minced on an iced petri dish. Minced tissue was placed 25 in an iced homogenizer (with 10 mL media) and disrupted with ten (10) strokes of a motorized Teflon pestle. The volume of homogenate was adjusted with the appropriate amount of media for each incubation set. Final incubation volume per inventive compound was 3 mL. Approximate protein concentration was 30 mg/mL. Incubation time, sampling rate and compound concentrations were the same as used for whole blood incubation, described in 30 Example 22, above. Lisofylline was incubated in the same conditions as the inventive compounds to demonstrate recovery of lisofylline in the system.

Figures 5 and 6 illustrate the hydrolysis of the various inventive compounds in the presence of human liver homogenates. As observed in the previous examples, the compound most resistant to hydrolysis is 1570R. A medium rate of hydrolysis was observed for 35 compound no. 4547R and rapid hydrolysis, in comparison to other compounds, was observed for compounds nos. 1515R, 4544R, 4545R and 4546R. The rapid hydrolysis of the inventive

compounds 4544 and 4545 by the liver homogenate illustrates their specific high hydrolytic potential in this tissue.

Example 25

As the data provided in examples 22 -24 were representative, predictive *in vitro* assays of conditions that a compound would experience upon oral administration, this example, showing *in vivo* results conducted in canine species, confirms that representative compounds of the inventive genus (specifically, inventive compounds nos. 4544, 4545 and 4576): 1) may increase the bioavailability of the compound (for systemic hydrolysis to the therapeutic lisofylline); 2) may maintain a higher ratio of concentration of lisofylline (the desired therapeutic compound) to concentration of the S enantiomer of lisofylline; and 3) may retain this ratio for a longer period of time.

Research has shown that the dog metabolizes lisofylline in a manner similar to humans; thus, this representative assay is predictive of results which would be seen in oral administration of the inventive compounds in humans.

Procedurally, stereochemical HPLC analyses were conducted as outlined in example 22 above. Treatment of dogs (in duplicate) with various compounds was accomplished by administration of a single oral dose (10 mg/kg) via a mouth tube. The test compounds were first dissolved into water and dosed as solutions. Venous blood draws were conducted at follows: predose, and then 5, 10, 20, 30, 60, 120 and 240 minutes post dose. The blood samples were centrifuged to obtain the plasma, which was drawn off and frozen at -70 °C until analyzed.

Figures 7-10 depict plasma levels of lisofylline and its corresponding S enantiomer in plasma of dogs that were treated with the inventive compound nos. 4544R, 4545R, 4576R and 4577R. From the data reported in these figures, the pharmacokinetics of lisofylline depend specifically on the chemical structure of a particular compound and its corresponding hydrolytic potential that was administered. In particular, the absolute amounts of target therapeutic, lisofylline, that are formed were dependent upon which specific compound was orally administered.

Using compound no. 4544R, the absolute canine blood levels of lisofylline are the most significant, at 1000 ng/mL. The level achieved with 4545R and 4577R represent intermediate blood levels at approximately 400 ng/mL. The lowest blood levels obtained for the compounds tested were 200 ng/mL for compound no. 4576R.

Due to the metabolic profile for lisofylline, an additional quantitative measure in evaluating the hydrolytic potential of the inventive compounds is a ratio of lisofylline to its corresponding S enantiomer (R:S ratio), which as confirmed by the data obtained also depends on the chemical entity administered and its structural chemistry.

In figure 8, the maximum concentrations of lisofylline and its S enantiomer are equal and appear about 0.25 hours after administration of inventive compound no. 4545. In figure 9, inventive compound no. 4576 results in a more significant and prolonged increase in the maximum concentration of the desired pharmaceutical lisofylline, as compared with its corresponding S enantiomer. The ratio of lisofylline to S enantiomer is approximately 1 at that point and drops thereafter. A comparison of figures 8 with 7, 9 and 10 illustrates that an R:S ratio substantially greater than unity can be achieved with compounds nos. 4544R, 4576R and 4577R; however, compound no. 4545R suggests that the corresponding S enantiomer is present in greater quantities than lisofylline.

An additional property shown exclusively by compound no. 4576R is an increased lisofylline half-life. This characteristic is best illustrated by comparing the slope of the elimination phase of lisofylline in figure 9 against either figure 7 or 8. Also, in analyzing these data, compound no. 4577R exhibits an interesting feature, involving an evident delay in reaching a maximum blood concentration (Tmax). Administering compound no. 4577R as described above delays Tmax 1 hour. However, Tmax occurs much earlier (within 0.5 hour) using other representative compounds.

These data suggest that specific compounds have greater hydrolytic potential based on their chemical structure and exhibit certain desirable characteristics in this predictive metabolic profile.